

Installing packages

```
In [5]: #much of this tutorial is derived from the scanpy tutorial here:
        #https://scanpy-tutorials.readthedocs.io/en/latest/pbmc3k.html

        #getting packages that you need for this analysis using the linux package manager pip
        !pip install scanpy #the package we will be using for the bulk of our ana
        lysis
        !pip3 install leidenalg #the algorithm we will use for clustering
```

Collecting scanpy

Downloading scanpy-1.8.1-py3-none-any.whl (2.0 MB)

2.0 MB 5.2 MB/s eta 0:00:01

Collecting tqdm

Downloading tqdm-4.62.3-py2.py3-none-any.whl (76 kB)

76 kB 6.6 MB/s eta 0:00:01

Collecting natsort

Downloading natsort-7.1.1-py3-none-any.whl (35 kB)

Collecting anndata>=0.7.4

Downloading anndata-0.7.6-py3-none-any.whl (127 kB)

127 kB 70.7 MB/s eta 0:00:01

Collecting sinfo

Downloading sinfo-0.3.4.tar.gz (24 kB)

Collecting importlib metadata>=0.7

Downloading importlib_metadata-4.8.1-py3-none-any.whl (17 kB)

Collecting scikit-learn>=0.22

Downloading scikit_learn-0.24.2-cp37-cp37m-manylinux2010_x86_64.whl (22.3 MB)

22.3 MB 50.0 MB/s eta 0:00:01

Collecting umap-learn>=0.3.10

Downloading umap-learn-0.5.1.tar.gz (80 kB)

80 kB 12.8 MB/s eta 0:00:01

Collecting packaging

Downloading packaging-21.0-py3-none-any.whl (40 kB)

40 kB 8.1 MB/s eta 0:00:01

Collecting scipy>=1.4

Downloading scipy-1.7.1-cp37-cp37m-manylinux_2_5_x86_64.manylinux1_x86_64.whl (28.5 MB)

28.5 MB 68.6 MB/s eta 0:00:01

Collecting pandas>=0.21

```
Downloading pandas-1.3.3-cp37-cp37m-manylinux_2_17_x86_64.manylinux2014_x86_64.whl (11.3 MB)
```

[illegible]

Collecting h5py>=2.10.0

Downloading h5py-3.4.0-cp37-cp37m-manylinux_2_12_x86_64.manylinux2010_x86_64.whl (4.1 MB)

4.1 MB 64.3 MB/s eta 0:00:01

Collecting seaborn

Downloading seaborn-0.11.2-py3-none-any.whl (292 kB)

292 kB 74.3 MB/s eta 0:00:01

Collecting statsmodels>=0.10.0rc2

```
Downloading statsmodels-0.13.0rc0-cp37-cp37m-manylinux_2_17_x86_64.manylinux2014_x86_64.whl (9.8 MB)
```

9.8 MB 27.8 MB/s eta 0:00:01

Collecting patsy

Downloading patsy-0.5.1-py2.py3-none-any.whl (231 kB)


```

Downloading threadpoolctl-2.2.0-py3-none-any.whl (12 kB)
Collecting pynndescent>=0.5
  Downloading pynndescent-0.5.4.tar.gz (1.1 MB)
    1.1 MB 64.4 MB/s eta 0:00:01
Collecting stdlib_list
  Downloading stdlib_list-0.8.0-py3-none-any.whl (63 kB)
    63 kB 3.2 MB/s eta 0:00:01
Collecting numexpr>=2.6.2
  Downloading numexpr-2.7.3-cp37-cp37m-manylinux2010_x86_64.whl (471 kB)
    471 kB 73.2 MB/s eta 0:00:01
Building wheels for collected packages: umap-learn, pynndescent, sinfo
  Building wheel for umap-learn (setup.py) ... done
  Created wheel for umap-learn: filename=umap_learn-0.5.1-py3-none-any.whl size=76566 sha256=7673e9c8b55e7a7a3bd626c57be341a79aacbaf35944058ecd6b01ae19955d4a
  Stored in directory: /home/jupyter/.cache/pip/wheels/01/e7/bb/347dc0e510803d7116a13d592b10cc68262da56a8eec4dd72f
  Building wheel for pynndescent (setup.py) ... done
  Created wheel for pynndescent: filename=pynndescent-0.5.4-py3-none-any.whl size=52372 sha256=94909d90b75379ca634c4d9930d188a8b7a977a0e648d9807e374b2228a3220d
  Stored in directory: /home/jupyter/.cache/pip/wheels/d0/5b/62/3401692ddad12324249c774c4b15ccb046946021e2b581c043
  Building wheel for sinfo (setup.py) ... done
  Created wheel for sinfo: filename=sinfo-0.3.4-py3-none-any.whl size=7899 sha256=cf86aa1e3999f749bcb7eed9177ca2071f6f9379573403af03c0fc211507a59
  Stored in directory: /home/jupyter/.cache/pip/wheels/68/ca/56/344d532fe53e855ccd6549795d370588ab8123907eecf4cf30
Successfully built umap-learn pynndescent sinfo
Installing collected packages: numpy, threadpoolctl, six, setuptools, scipy, llvmlite, joblib, zipp, typing-extensions, scikit-learn, pytz, python-dateutil, pyparsing, pillow, numba, kiwisolver, cycycler, cached-property, xlrd, stdlib-list, pynndescent, patsy, pandas, packaging, numexpr, natsort, matplotlib, importlib-metadata, h5py, umap-learn, tqdm, tables, statsmodels, sinfo, seaborn, networkx, anndata, scanpy
ERROR: pip's dependency resolver does not currently take into account all the packages that are installed. This behaviour is the source of the following dependency conflicts.
tensorflow 2.4.2 requires h5py~=2.10.0, but you have h5py 3.4.0 which is incompatible.
tensorflow 2.4.2 requires numpy~=1.19.2, but you have numpy 1.20.3 which is incompatible.
tensorflow 2.4.2 requires six~=1.15.0, but you have six 1.16.0 which is incompatible.
tensorflow 2.4.2 requires typing-extensions~=3.7.4, but you have typing-extensions 3.10.0.2 which is incompatible.
jupyterlab-git 0.11.0 requires nbdtimes<2.0.0, >=1.1.0, but you have nbdtimes 3.1.0 which is incompatible.
Successfully installed anndata-0.7.6 cached-property-1.5.2 cycycler-0.10.0 h5py-3.4.0 importlib-metadata-4.8.1 joblib-1.0.1 kiwisolver-1.3.2 llvmlite-0.37.0 matplotlib-3.4.3 natsort-7.1.1 networkx-2.6.3 numba-0.54.0 numexpr-2.7.3 numpy-1.20.3 packaging-21.0 pandas-1.3.3 patsy-0.5.1 pillow-8.3.2 pynndescent-0.5.4 pyparsing-2.4.7 python-dateutil-2.8.2 pytz-2021.1 scanpy-1.8.1 scikit-learn-0.24.2 scipy-1.7.1 seaborn-0.11.2 setuptools-58.0.4 sinfo-0.3.4 six-1.16.0 statsmodels-0.13.0rc0 stdlib-list-0.8.0 tables-3.6.1 threadpoolctl-2.2.0 tqdm-4.62.3 typing-extensions-3.10.0.2 umap-learn-0.5.1 xlrd-1.2.0 zipp-3.5.0
WARNING: You are using pip version 21.1.3; however, version 21.2.4 is available.
You should consider upgrading via the '/opt/conda/bin/python3.7 -m pip install --upgrade pip' command.
Collecting leidenalg
  Downloading leidenalg-0.8.7-cp37-cp37m-manylinux2010_x86_64.whl (1.4 MB)
    1.4 MB 5.1 MB/s eta 0:00:01
Collecting python-igraph>=0.9.0
  Downloading python_igraph-0.9.6-cp37-cp37m-manylinux2010_x86_64.whl (3.2 MB)
    3.2 MB 62.7 MB/s eta 0:00:01
Collecting texttable>=1.6.2
  Downloading texttable-1.6.4-py2.py3-none-any.whl (10 kB)
Installing collected packages: texttable, python-igraph, leidenalg
Successfully installed leidenalg-0.8.7 python-igraph-0.9.6 texttable-1.6.4

```

WARNING: Target directory /home/jupyter/notebooks/packages/__pycache__ already exists. Specify --upgrade to force replacement.

WARNING: Target directory /home/jupyter/notebooks/packages/bin already exists. Specify --upgrade to force replacement.

WARNING: You are using pip version 21.1.3; however, version 21.2.4 is available.

You should consider upgrading via the '/opt/conda/bin/python3.7 -m pip install --upgrade pip' command.

Getting the single cell data from a google bucket

```
In [2]: #going and getting the h5ad file, processed through cumulus by Nathan Tucker's comp bio person
#cumulus workspace available here: https://app.terra.bio/#workspaces/kco-tech/Cumulus

#this is from a human aorta data set, we got this dataset from the human cell atlas:
#https://data.humancellatlas.org/explore/projects/07073c12-8006-4710-a00b-23abdb814904

!mkdir data #making a storage directory to put the data!

#here we actually go and get the data. gsutil is a google cloud utility to copy data from
#"buckets" into our local disk space. Then we will be able to ingest and manipulate the data
!gsutil cp gs://fc-9c9cf8d0-aafd-47c2-ab1b-4e0f20ab200b/aortic_adata_full_li_lemaire.h5ad data/
```

Copying gs://fc-9c9cf8d0-aafd-47c2-ab1b-4e0f20ab200b/aortic_adata_full_li_lemaire.h5ad...
/[1 files][4.7 GiB/ 4.7 GiB] 59.2 MiB/s
Operation completed over 1 objects/4.7 GiB.

```
In [42]: #listing the contents of our data directory
!ls data/
```

aortic_adata_full_li_lemaire.h5ad

Setting up the python environment

```
In [43]: #importing necessary python packages
import numpy as np
import pandas as pd
import scanpy as sc

#setting parameters for scanpy
sc.settings.verbosity = 3 # verbosity: errors (0), warnings (1), info (2), hints (3)
sc.logging.print_versions() #version checking.....
sc.settings.set_figure_params(dpi=80)
```

WARNING: If you miss a compact list, please try `print_header`!

The `sinfo` package has changed name and is now called `session_info` to become more discoverable and self-explanatory. The `sinfo` PyPI package will be kept around to avoid breaking old installs and you can downgrade to 0.3.2 if you want to use it without seeing this message. For the latest features and bug fixes, please install `session_info` instead. The usage and defaults also changed slightly, so please review the latest README at https://gitlab.com/joelostblom/session_info.

anndata 0.7.6

scanpy	1.8.1	
sinfo	0.3.4	

PIL	8.2.0	
attr	21.2.0	
backcall	0.2.0	
beta_ufunc	NA	
binom_ufunc	NA	
bottleneck	1.3.2	
brotili	NA	
cached_property	1.5.2	
cachetools	4.2.2	
caip_notebooks_serverextension	NA	
certifi	2021.05.30	
cffi	1.14.5	
chardet	4.0.0	
cloudpickle	1.6.0	
colorama	0.4.4	
cryptography	3.4.7	
cycler	0.10.0	
cython_runtime	NA	
dateutil	2.8.1	
decorator	5.0.9	
google	NA	
greenlet	1.1.0	
grpc	1.32.0	
h5py	3.4.0	
idna	2.10	
igraph	0.9.6	
ipykernel	5.5.5	
ipython_genutils	0.2.0	
ipywidgets	7.6.3	
jedi	0.18.0	
jinja2	3.0.1	
joblib	1.0.1	
jsonschema	3.2.0	
jwt	2.1.0	
kiwisolver	1.3.1	
leidenalg	0.8.7	
llvmlite	0.37.0	
markupsafe	2.0.1	
matplotlib	3.4.2	
matplotlib_inline	NA	
mpl_toolkits	NA	
natsort	7.1.1	
nbformat	5.1.3	
nbinom_ufunc	NA	
numba	0.54.0	
numexpr	2.7.3	
numpy	1.19.5	
packaging	20.9	
pandas	1.2.4	
parso	0.8.2	
pexpect	4.8.0	
pickleshare	0.7.5	
pkg_resources	NA	
prettytable	2.1.0	

```
prometheus_client      NA
prompt_toolkit          3.0.19
proto                   NA
psutil                  5.8.0
ptyprocess              0.7.0
pvectorc                NA
pyarrow                 4.0.1
pygments                2.9.0
pyparsing               2.4.7
pysistent              NA
pytz                    2021.1
requests                2.25.1
scipy                   1.7.1
seaborn                 0.11.2
send2trash              NA
simplejson               3.17.2
six                     1.15.0
sklearn                 0.24.2
socks                   1.7.1
sql                     NA
sqlalchemy              1.4.18
sqlparse                0.4.1
statsmodels             0.13.0rc0
storemagic              NA
tables                  3.6.1
terminado               0.10.1
texttable               1.6.4
tornado                 6.1
tqdm                    4.61.1
traitlets               5.0.5
typing_extensions       NA
uritemplate             3.0.1
urllib3                 1.26.5
wcwidth                 0.2.5
yaml                    5.4.1
zipp                    NA
zmq                     22.1.0
```

```
IPython                 7.24.1
jupyter_client          6.1.12
jupyter_core            4.7.1
jupyterlab              1.2.16
notebook                 6.4.0
```

Python 3.7.10 | packaged by conda-forge | (default, Feb 19 2021, 16:07:37) [GCC 9.3.0]

Linux-5.4.104+-x86_64-with-debian-buster-sid

8 logical CPU cores, x86_64

Session information updated at 2021-09-21 16:35

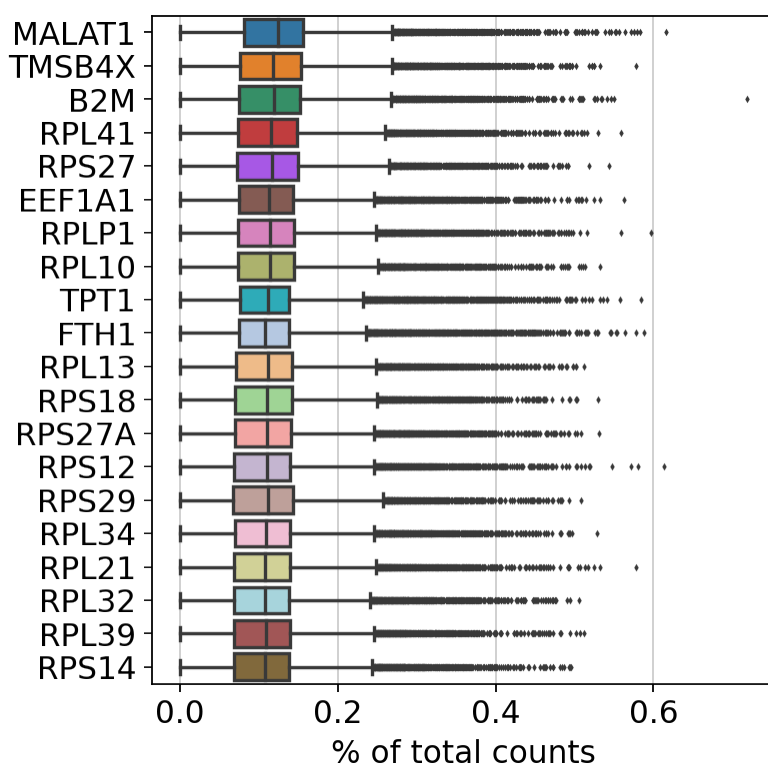
Reading in & processing the single cell aorta data h5ad file

```
In [44]: #reading in the data using the scanpy function sc.read
adata = sc.read("data/aortic_adata_full_li_lemaire.h5ad") #this file is huge, will take a minute to read in (4.7Gb)
adata #this will show properties of the adata object, a very useful first step
```

```
Out[44]: AnnData object with n_obs × n_vars = 27225 × 23318
  obs: 'Sample', 'Treatment', 'batch', 'n_genes', 'n_genes_by_counts', 'total_counts', 'total_counts_mt', 'pct_counts_mt', 'leiden'
  var: 'mt', 'gene_ids', 'n_cells_by_counts', 'mean_counts', 'pct_dropout_by_counts', 'total_counts', 'highly_variable', 'means', 'dispersions', 'dispersions_norm'
  uns: 'Sample_colors', 'Treatment_colors', 'hvg', 'leiden', 'leiden_colors', 'neighbors', 'pca', 'umap'
  obsm: 'X_pca', 'X_umap'
```

```
In [45]: #Show those genes that yield the highest fraction of counts in each single cell, across all cells.
sc.pl.highest_expr_genes(adata, n_top=20, )
```

normalizing counts per cell
finished (0:00:01)



Some filtering of the data

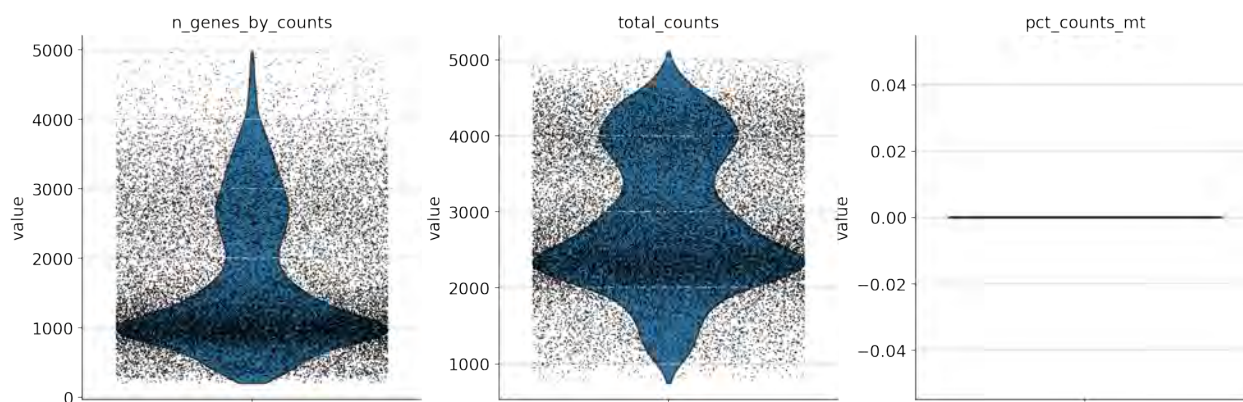
```
In [46]: #basic filtering:
#cells need to have at least 200 genes expressed
#genes need to be expressed in at least 3 cells

sc.pp.filter_cells(adata, min_genes=200)
sc.pp.filter_genes(adata, min_cells=3)
```

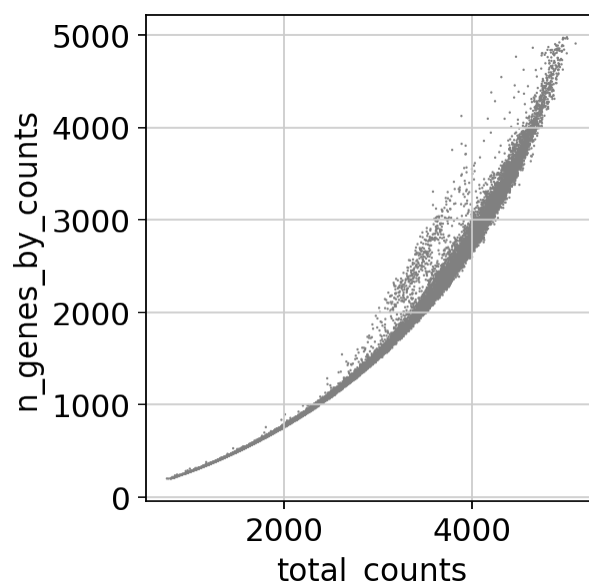
filtered out 1 cells that have less than 200 genes expressed
filtered out 1675 genes that are detected in less than 3 cells

```
In [47]: adata.var['mt'] = adata.var_names.str.startswith('MT-') # annotate the group of mi
         tochondrial genes as 'mt'
         sc.pp.calculate_qc_metrics(adata, qc_vars=['mt'], percent_top=None, log1p=
         False, inplace=True)
```

```
In [48]: #plot the result of the previous calculation
         sc.pl.violin(adata, ['n_genes_by_counts', 'total_counts', 'pct_counts_mt'],
         ,
         jitter=0.4, multi_panel=True)
```



```
In [49]: #plot genes by counts against total counts per cell
         sc.pl.scatter(adata, x='total_counts', y='n_genes_by_counts')
```



```
In [50]: #total-count normalize (library-size correct) the data matrix to 10,000 reads per cell,
         #so that counts become comparable among cells.
         sc.pp.normalize_total(adata, target_sum=1e4)
```

normalizing counts per cell
finished (0:00:00)

```
In [51]: #log transform the data
         sc.pp.log1p(adata)
```

```
In [52]: #identify highly variable genes
```



```
sc.pp.highly_variable_genes(adata, min_mean=0.0125, max_mean=3, min_disp=0.5)
```

extracting highly variable genes

finished (0:00:06)

--> added

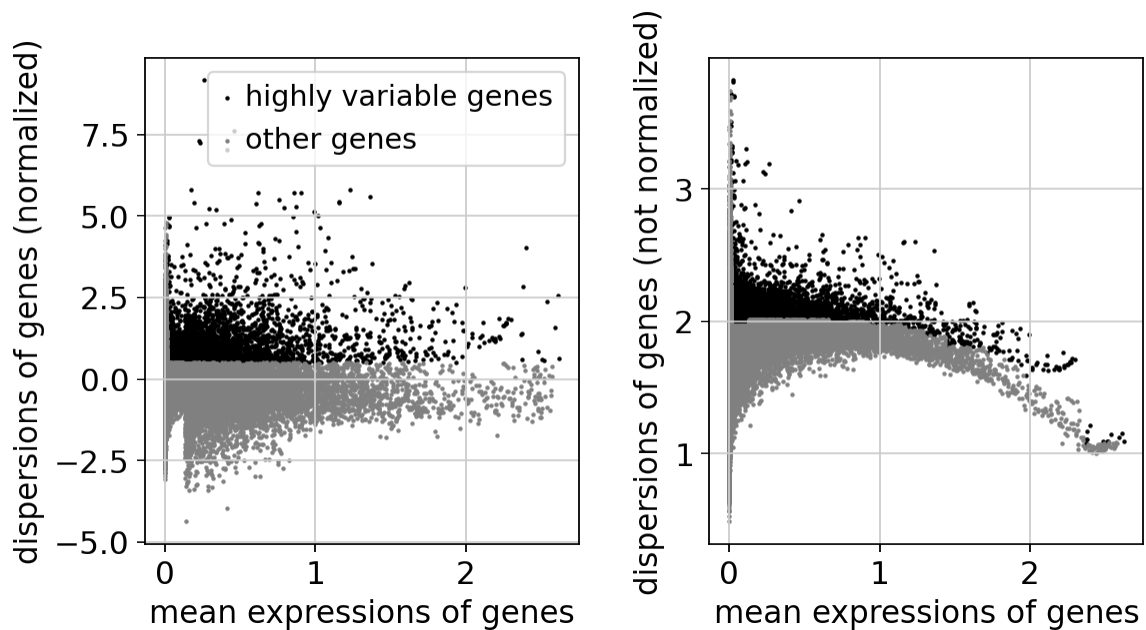
'highly_variable', boolean vector (adata.var)

'means', float vector (adata.var)

'dispersions', float vector (adata.var)

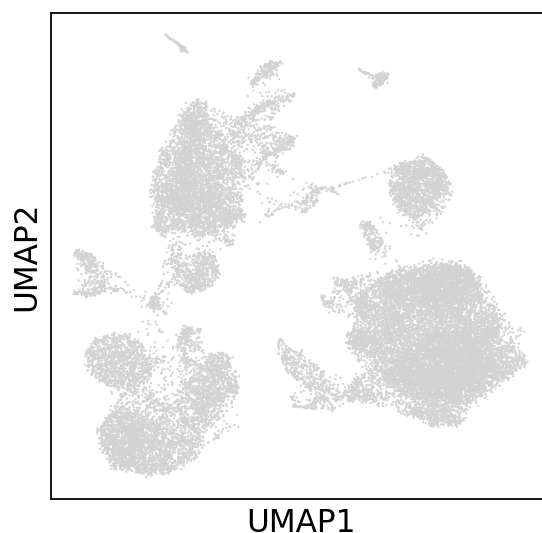
'dispersions_norm', float vector (adata.var)

```
In [53]: #plot the highly variable genes
sc.pl.highly_variable_genes(adata)
```

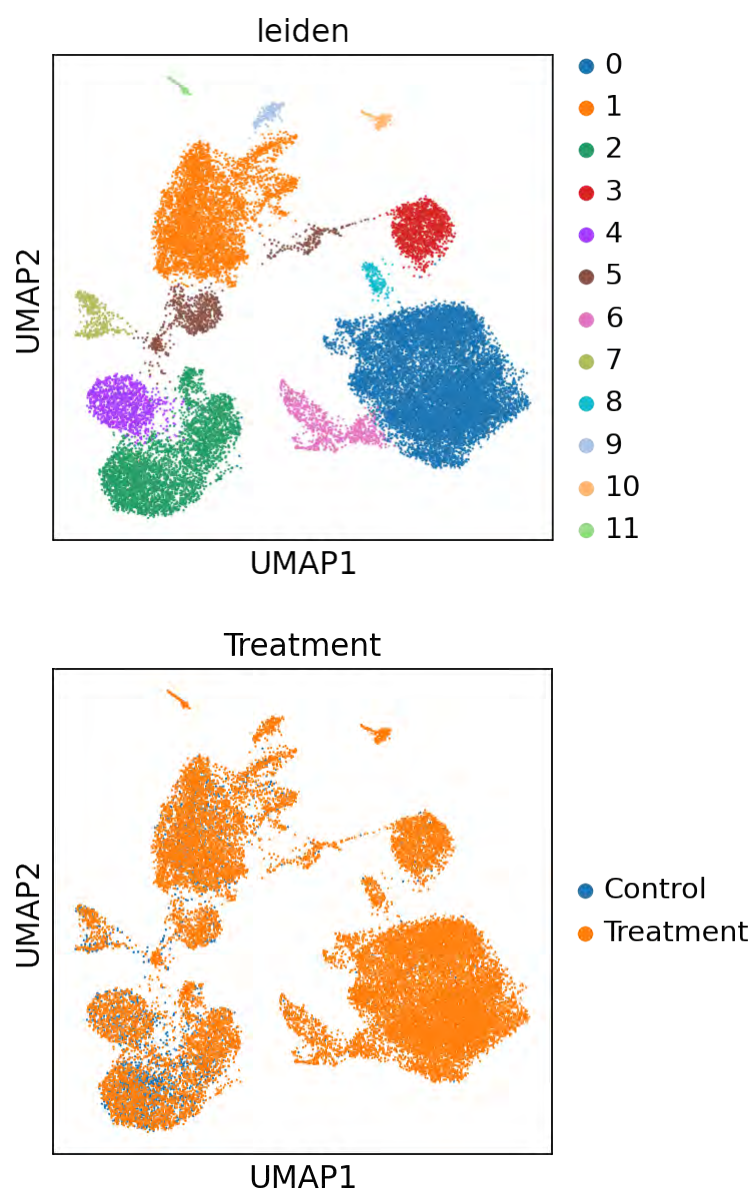


UMAPs and marker genes

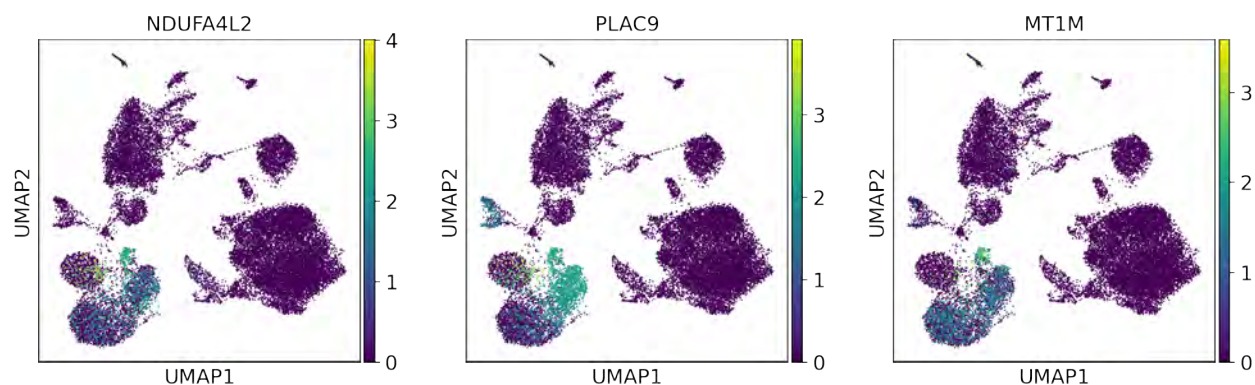
```
In [54]: #plot a basic UMAP
sc.pl.umap(adata)
```



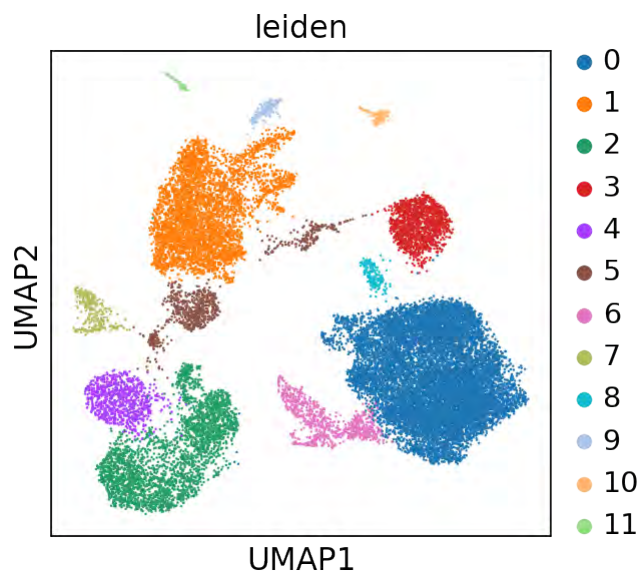
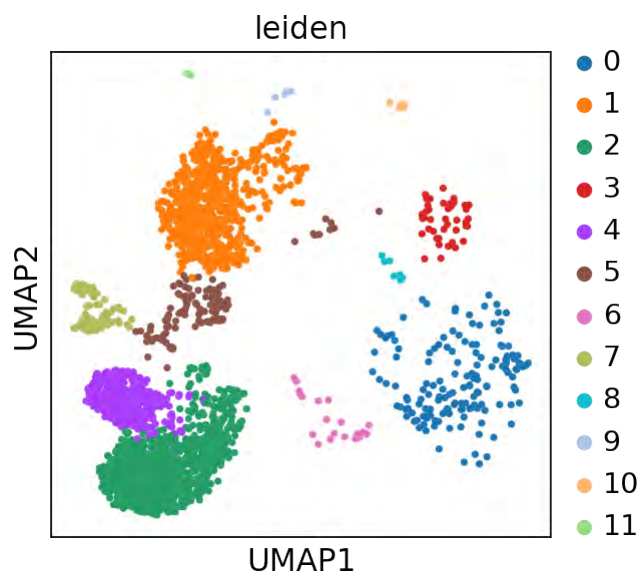
```
In [55]: #and plot the UMAP using louvain1.0 coloring scheme:
sc.pl.umap(adata, color=['leiden'])
sc.pl.umap(adata, color=['Treatment'])
```



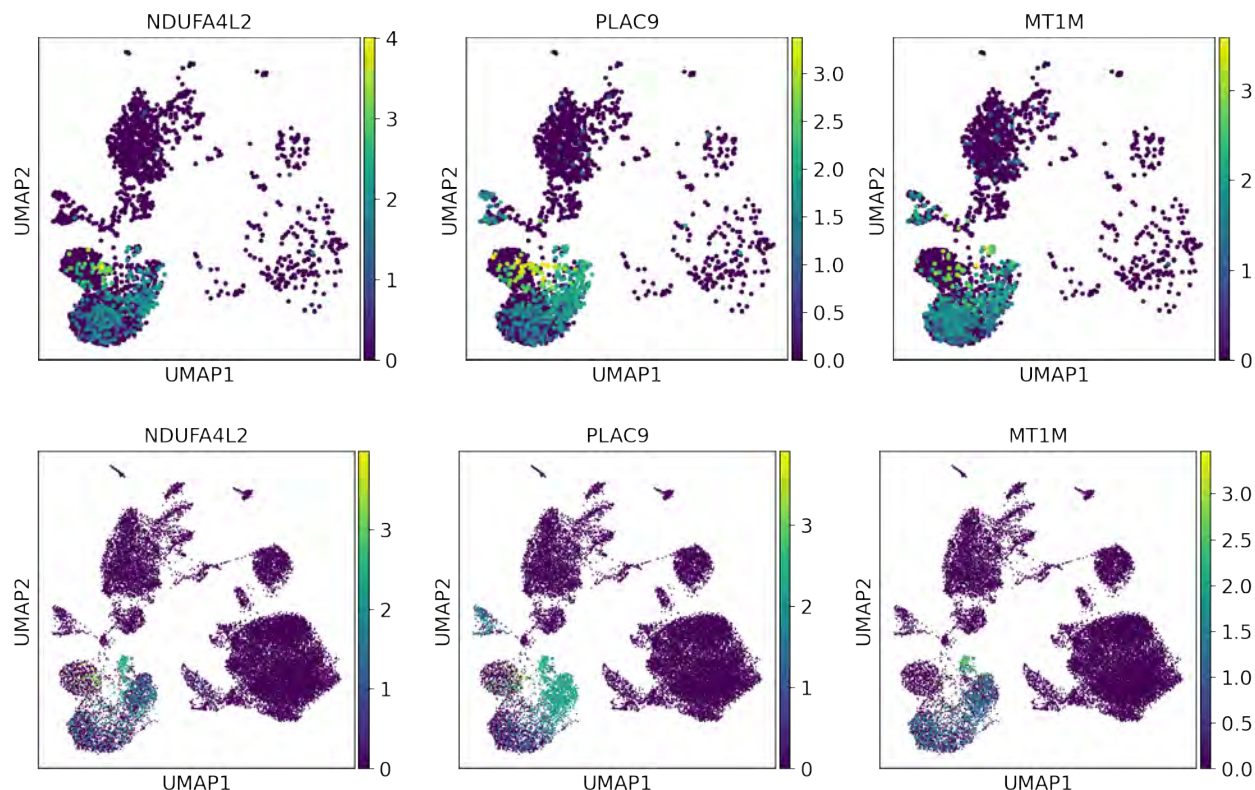
```
In [56]: #plot the UMAP coloring for a few genes of interest
#note: replace the text below with your specific genes of interest
sc.pl.umap(adata, color=['NDUFA4L2', 'PLAC9', 'MT1M'])
```



```
In [57]: #plotting leiden clustering for the Control and the Treatment (Aortic aneurysm)
sc.pl.umap(adata[adata.obs['Treatment'].isin(['Control'])], color='leiden')
sc.pl.umap(adata[adata.obs['Treatment'].isin(['Treatment'])], color='leiden')
```



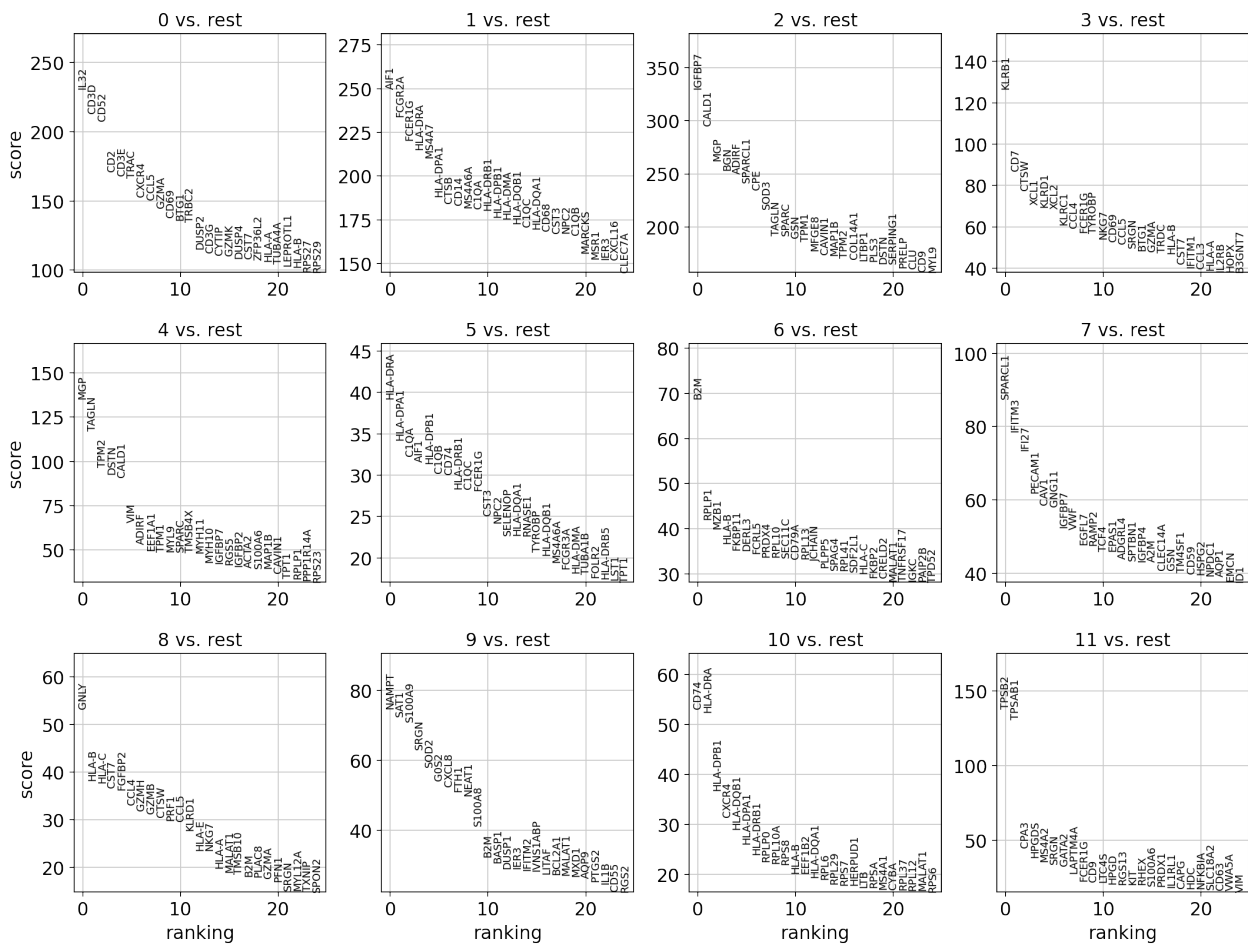
```
In [58]: #plotting genes of interest for the Control and the Treatment (Aortic aneurysm) for genes of interest
sc.pl.umap(adata[adata.obs['Treatment'].isin(['Control'])], color=['NDUFA4L2', 'PLAC9', 'MT1M'])
sc.pl.umap(adata[adata.obs['Treatment'].isin(['Treatment'])], color=['NDUFA4L2', 'PLAC9', 'MT1M'])
```



```
In [59]: #identifying marker genes using leiden clusters and a t-test
sc.tl.rank_genes_groups(adata, 'leiden', method='t-test')
sc.pl.rank_genes_groups(adata, n_genes=25, sharey=False)
```

ranking genes

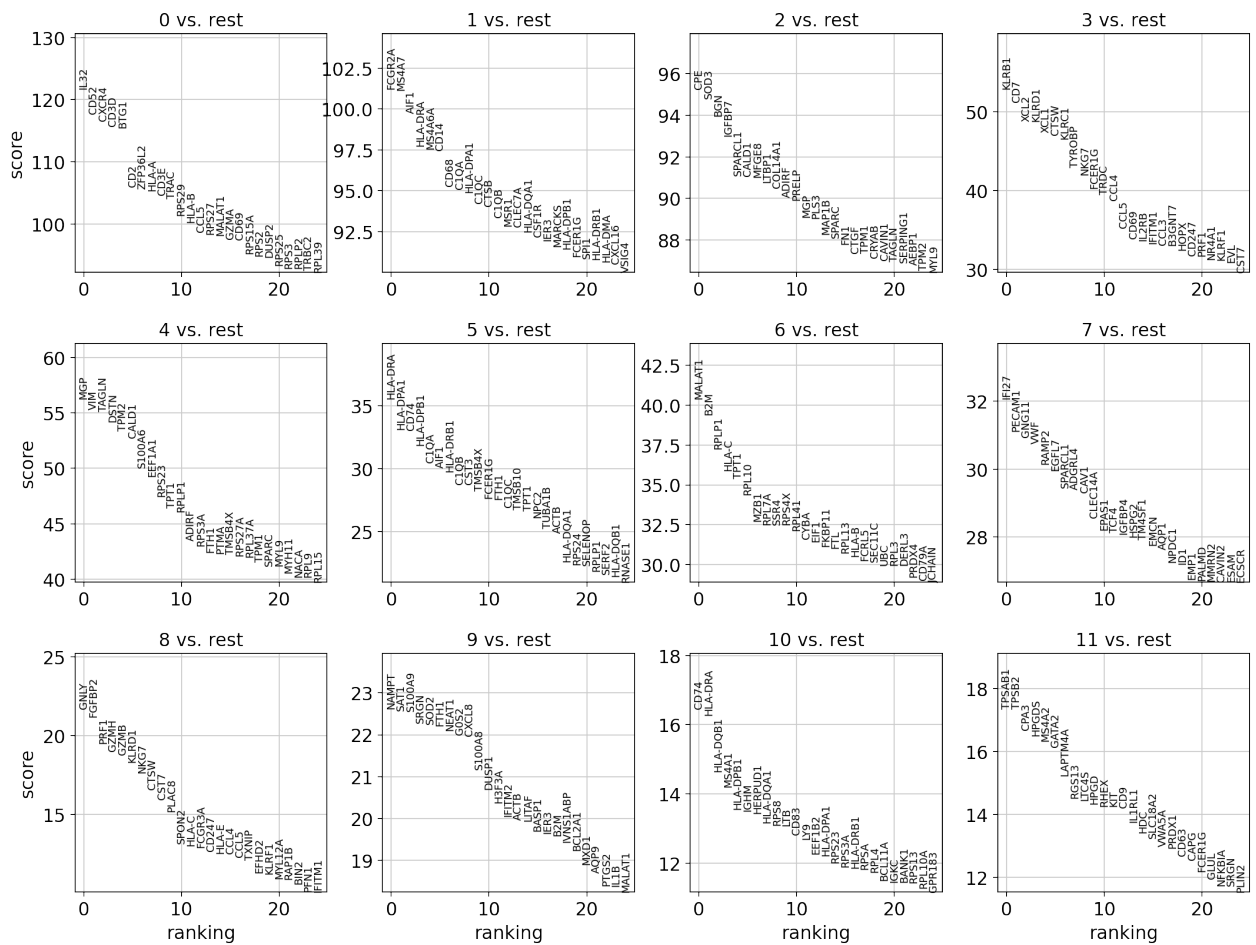
```
finished: added to `uns['rank_genes_groups']`
'names', sorted np.recarray to be indexed by group ids
'scores', sorted np.recarray to be indexed by group ids
'logfoldchanges', sorted np.recarray to be indexed by group ids
'pvals', sorted np.recarray to be indexed by group ids
'pvals_adj', sorted np.recarray to be indexed by group ids (0:00:42)
```



```
In [60]: #write this out to a file
adata.write('leiden_ttest_markers.txt')
```

```
In [61]: #identifying marker genes using wilcoxon rank-sum test
sc.tl.rank_genes_groups(adata, 'leiden', method='wilcoxon')
sc.pl.rank_genes_groups(adata, n_genes=25, sharey=False)
```

ranking genes
finished: added to `uns['rank_genes_groups']`
'names', sorted np.ndarray to be indexed by group ids
'scores', sorted np.ndarray to be indexed by group ids
'logfoldchanges', sorted np.ndarray to be indexed by group ids
'pvals', sorted np.ndarray to be indexed by group ids
'pvals_adj', sorted np.ndarray to be indexed by group ids (0:01:27)



In [62]: *#write this out to a file*

```
adata.write('leiden_wilcoxon_markers.txt')
```

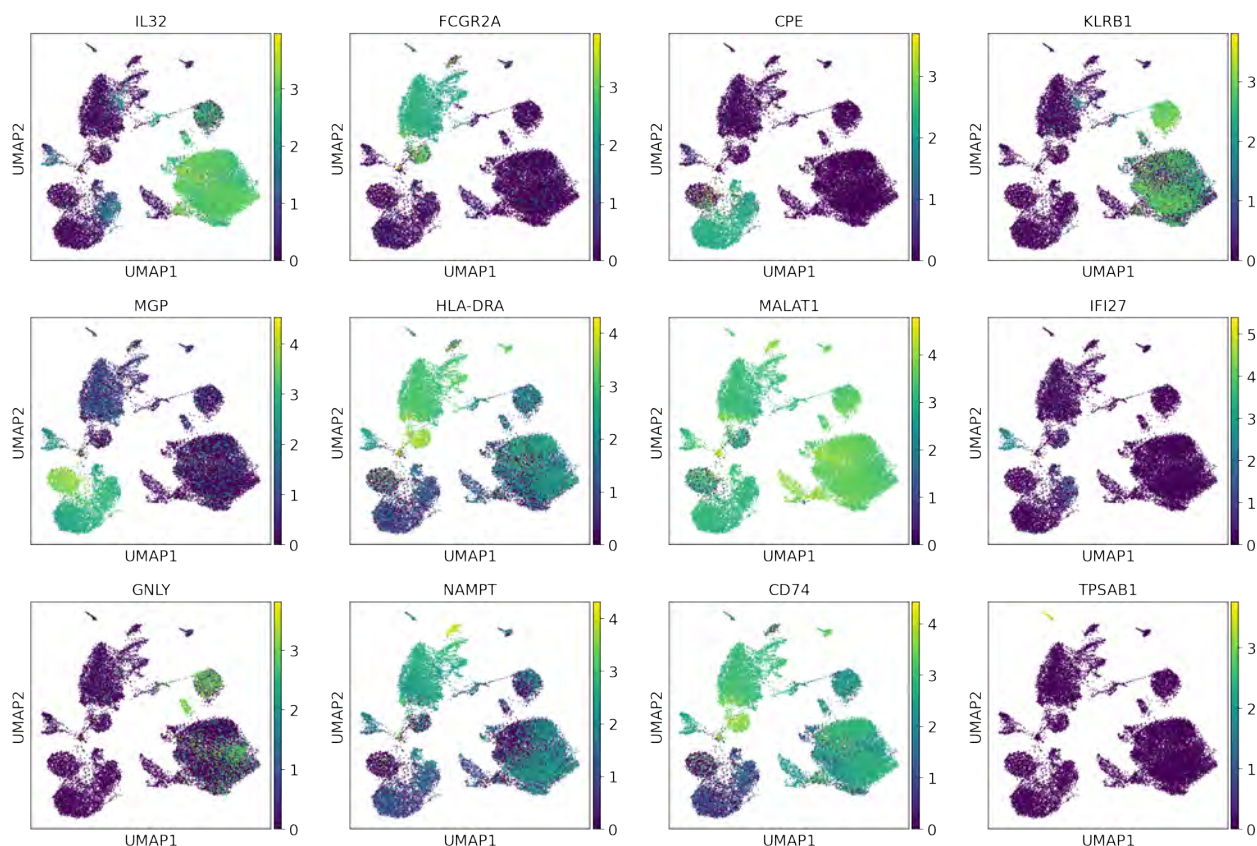
In [63]: *#defining the first ranked marker gene from wilcoxon rank sum testing*

```
marker_genes = ['IL32', 'FCGR2A', 'CPE', 'KLRB1', 'MGP', 'HLA-DRA', 'MALAT1', 'IFI27', 'GNLY', 'NAMPT', 'CD74', 'TPSAB1']
```

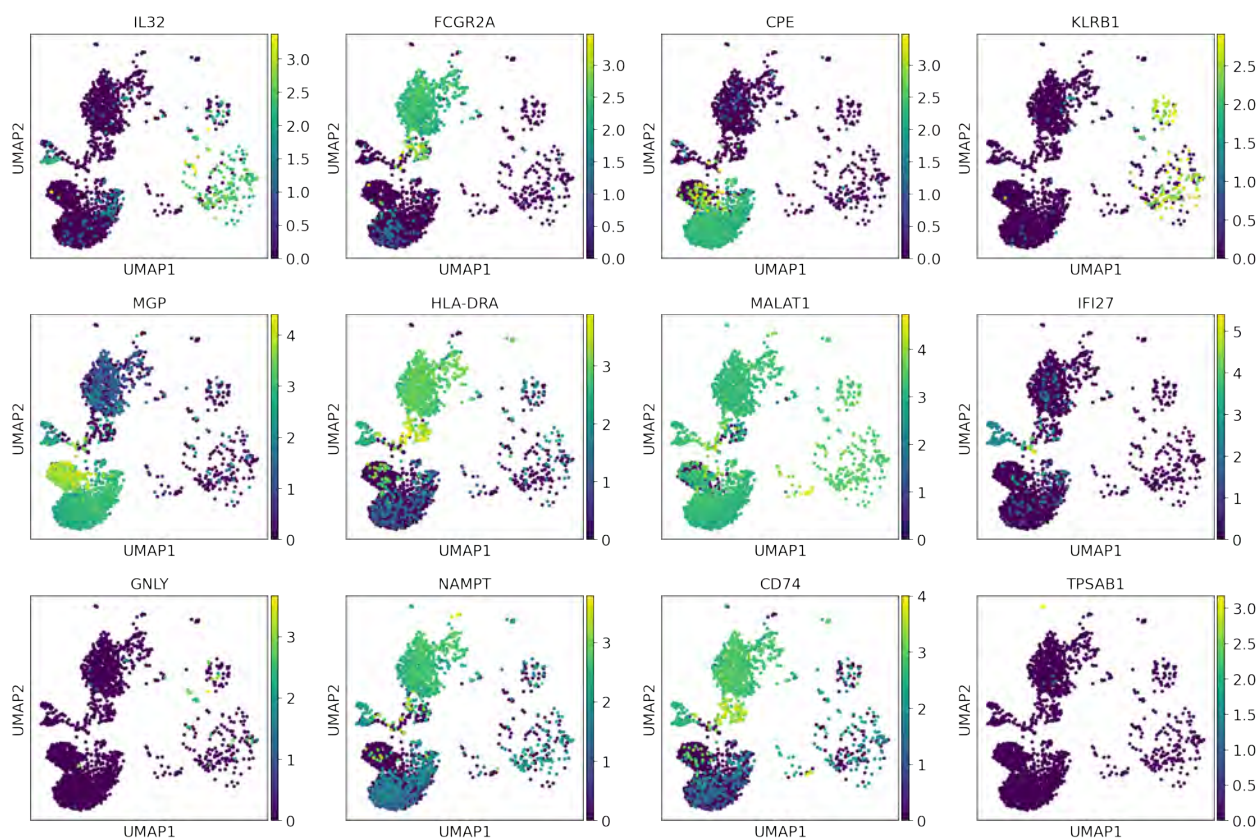
Examining the marker genes in the dataset

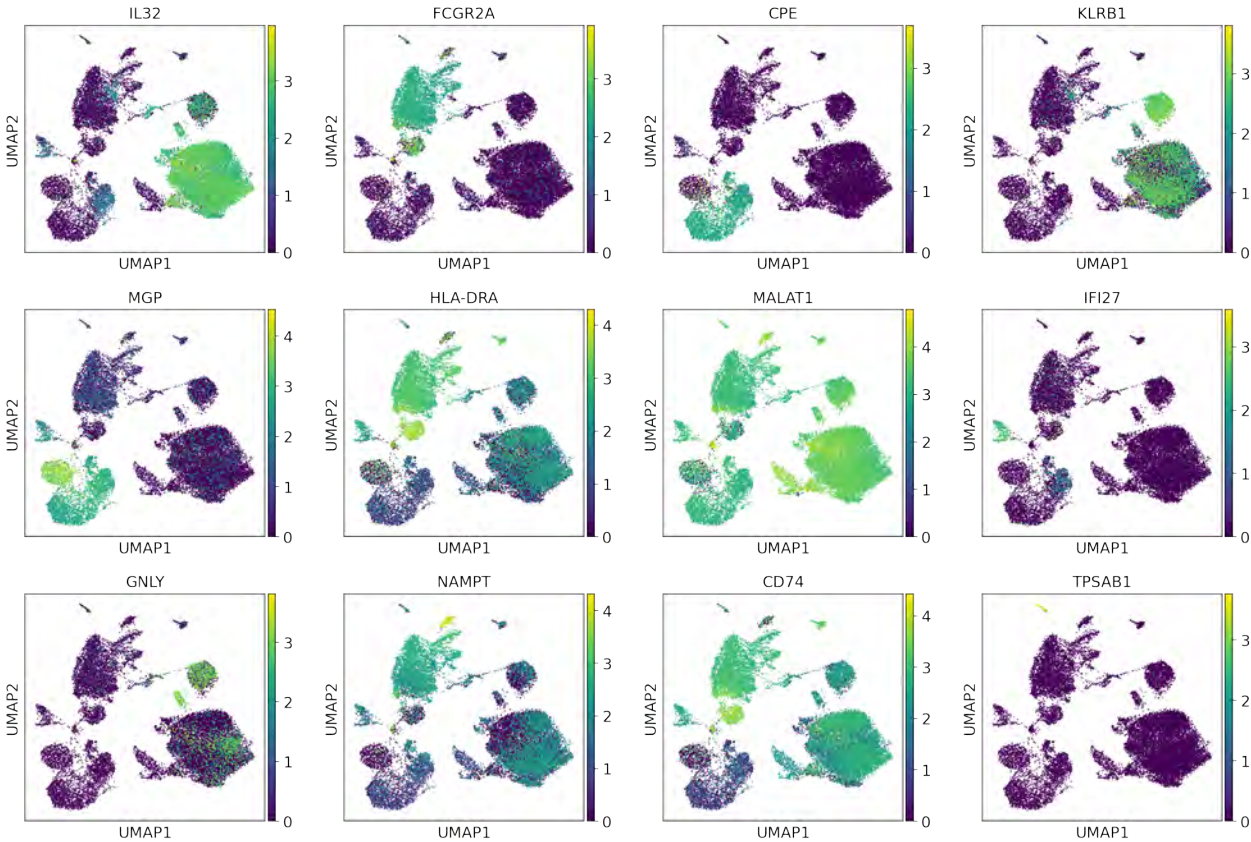
In [64]: *#plotting a umap of these marker genes*

```
sc.pl.umap(adata, color=marker_genes)
```

```
In [65]: #and doing this with respect to treatment/control cells
sc.pl.umap(adata[adata.obs['Treatment'].isin(['Control'])], color=marker_genes)
sc.pl.umap(adata[adata.obs['Treatment'].isin(['Treatment'])], color=marker_genes)
```





```
In [66]: #show the top 5 markers for each cluster!
pd.DataFrame(adata.uns['rank_genes_groups']['names']).head(5)
```

Out[66]:

	0	1	2	3	4	5	6	7	8	9	10	11
0	IL32	FCGR2A	CPE	KLRB1	MGP	HLA-DRA	MALAT1	IFI27	GNLY	NAMPT	CD74	TPSAB1
1	CD52	MS4A7	SOD3	CD7	VIM	HLA-DPA1	B2M	PECAM1	FGFBP2	SAT1	HLA-DRA	TPSB2
2	CXCR4	AIF1	BGN	XCL2	TAGLN	CD74	RPLP1	GNG11	PRF1	S100A9	HLA-DQB1	CPAC1
3	CD3D	HLA-DRA	IGFBP7	KLRD1	DSTN	HLA-DPB1	HLA-C	WWF	GZMH	SRGN	MS4A1	HPGD5
4	BTG1	MS4A6A	SPARCL1	XCL1	TPM2	C1QA	TPT1	RAMP2	GZMB	SOD2	HLA-DPB1	MS4A6

```
In [67]: #make a more detailed table, including p-values
result = adata.uns['rank_genes_groups']
groups = result['names'].dtype.names
pd.DataFrame(
    {group + '_' + key[:1]: result[key][group]
     for group in groups for key in ['names', 'pvals']}).head(5)
```

Out[67]:

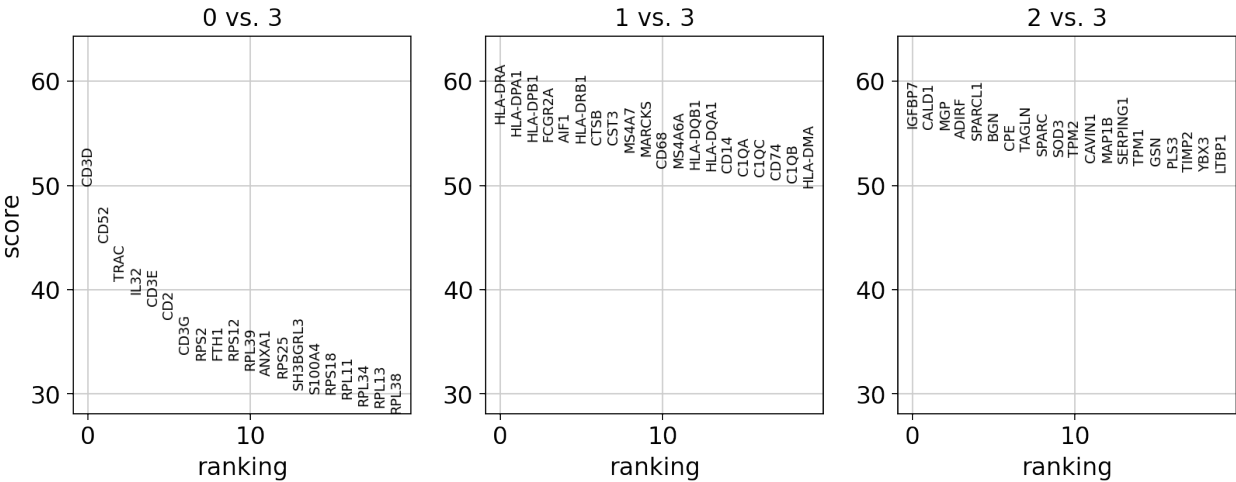
	0_n	0_p	1_n	1_p	2_n	2_p	3_n	3_p	4_n	4_p	...	7_n	7_p	8_n
0	IL32	0.0	FCGR2A	0.0	CPE	0.0	KLRB1	0.0	MGP	0.0	...	IFI27	9.019634e-226	GN

1	CD52	0.0	MS4A7	0.0	SOD3	0.0	CD7	0.0	VIM	0.0	...	PECAM1	1.612090e-212	FGFBP7
2	CXCR4	0.0	AIF1	0.0	BGN	0.0	XCL2	0.0	TAGLN	0.0	...	GNG11	3.551387e-210	PRKRA
3	CD3D	0.0	HLA-DRA	0.0	IGFBP7	0.0	KLRD1	0.0	DSTN	0.0	...	WWF	8.100193e-208	GZM1
4	BTG1	0.0	MS4A6A	0.0	SPARCL1	0.0	XCL1	0.0	TPM2	0.0	...	RAMP2	8.744787e-200	GZM1

5 rows × 24 columns

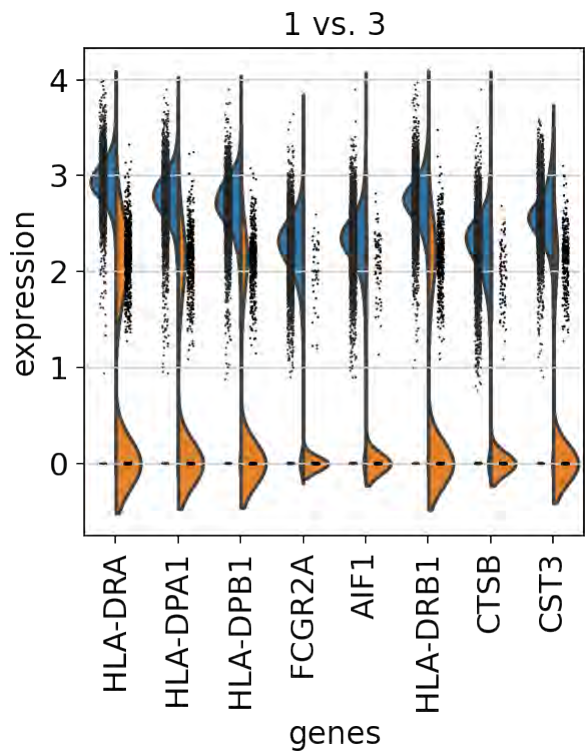
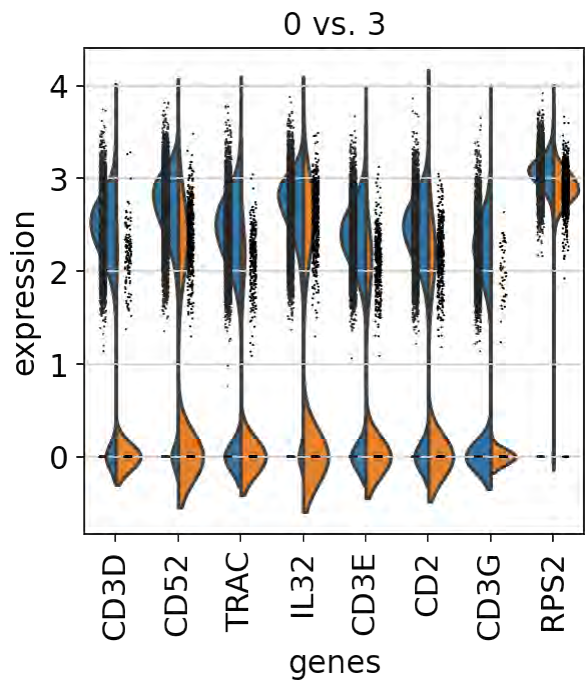
```
In [68]: #comparing two clusters (0 and 1)
sc.tl.rank_genes_groups(adata, 'leiden', groups=['0','1','2'], reference='3', method='wilcoxon')
sc.pl.rank_genes_groups(adata, groups=['0','1','2'], n_genes=20)
```

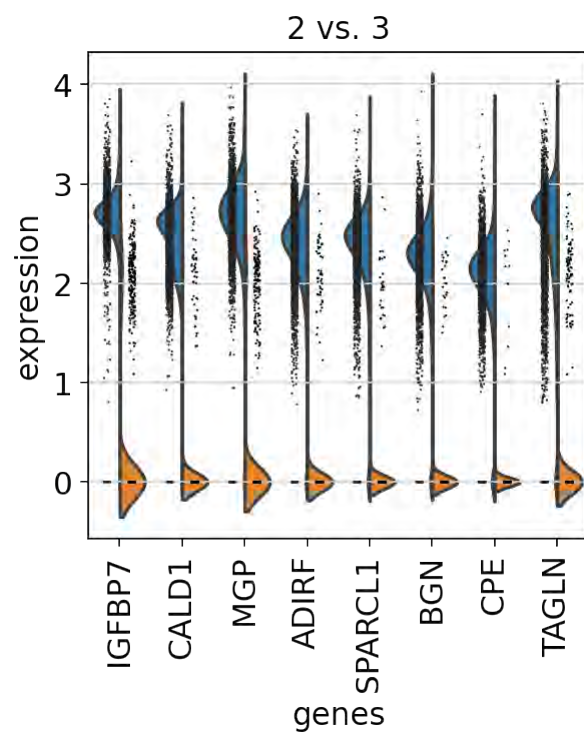
ranking genes
finished: added to `uns['rank_genes_groups']`
'names', sorted np.recarray to be indexed by group ids
'scores', sorted np.recarray to be indexed by group ids
'logfoldchanges', sorted np.recarray to be indexed by group ids
'pvals', sorted np.recarray to be indexed by group ids
'pvals_adj', sorted np.recarray to be indexed by group ids (0:00:44)



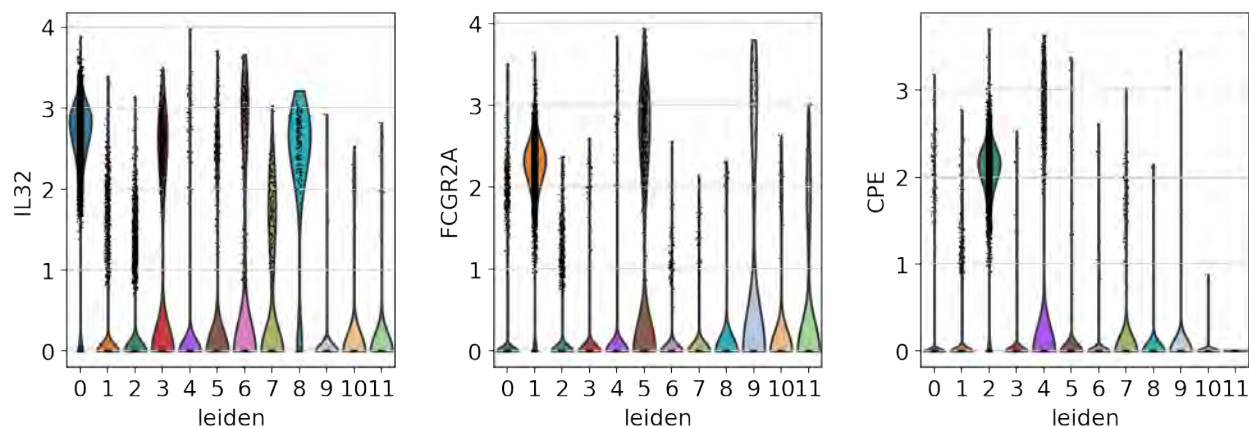
```
In [69]: #If we want a more detailed view for a certain group, use sc.pl.rank_genes_groups_violin.
sc.pl.rank_genes_groups_violin(adata, groups=['0','1','2'], n_genes=8)

#note that you can replace the "groups" and "n_genes" variables as you please here...
```





```
In [70]: #comparing different genes across the groups
sc.pl.violin(adata, ['IL32', 'FCGR2A', 'CPE'], groupby='leiden')
```

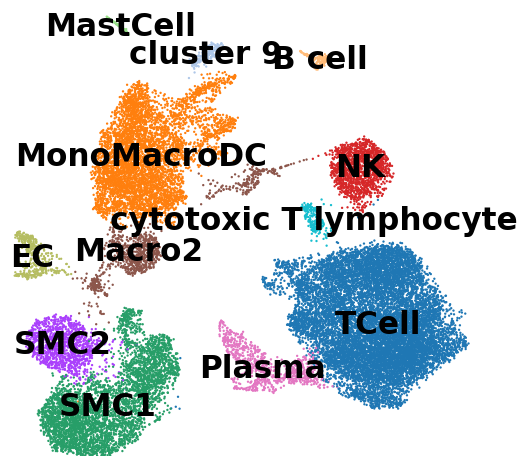


```
In [72]: #naming the clusters by their putative identities
#see https://www.ahajournals.org/doi/10.1161/CIRCULATIONAHA.120.046528
```

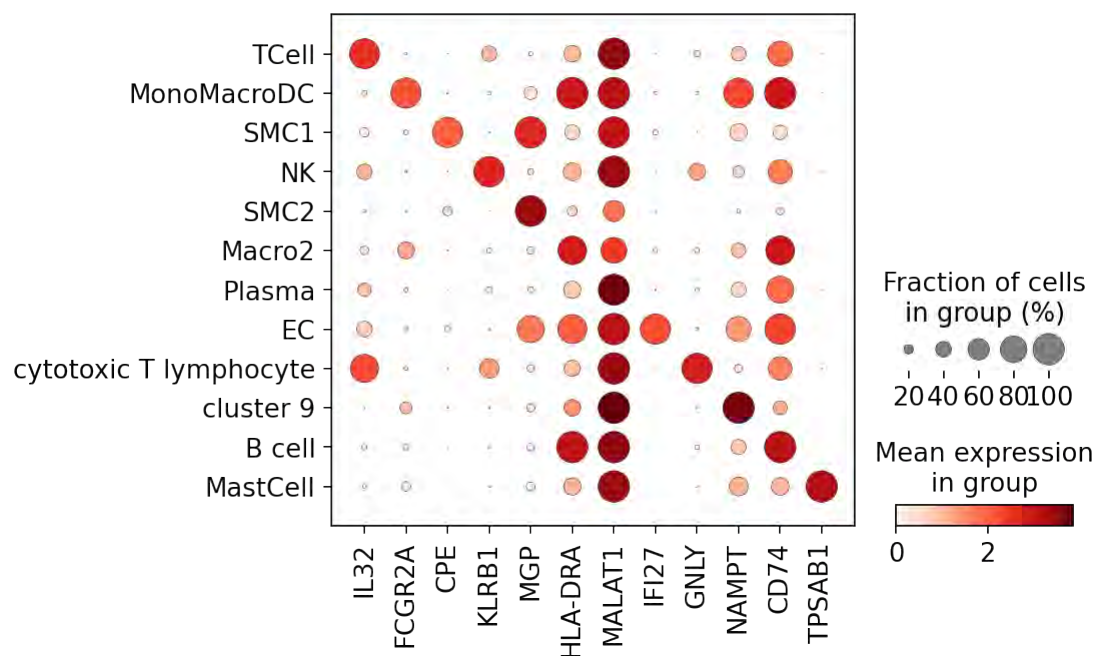
```
new_cluster_names = [
    'TCell',
    'MonoMacroDC',
    'SMC1',
    'NK',
    'SMC2',
    'Macro2',
    'Plasma',
    'EC',
    'cytotoxic T lymphocyte',
    'cluster 9',
    'B cell',
    'MastCell']
adata.rename_categories('leiden', new_cluster_names)
```

Omitting rank_genes_groups/names as old categories do not match.
 Omitting rank_genes_groups/scores as old categories do not match.
 Omitting rank_genes_groups/pvals as old categories do not match.
 Omitting rank_genes_groups/pvals_adj as old categories do not match.
 Omitting rank_genes_groups/logfoldchanges as old categories do not match.

```
In [73]: #making a umap with cluster names
sc.pl.umap(adata, color='leiden', legend_loc='on data', title='', frameon=
False)
```

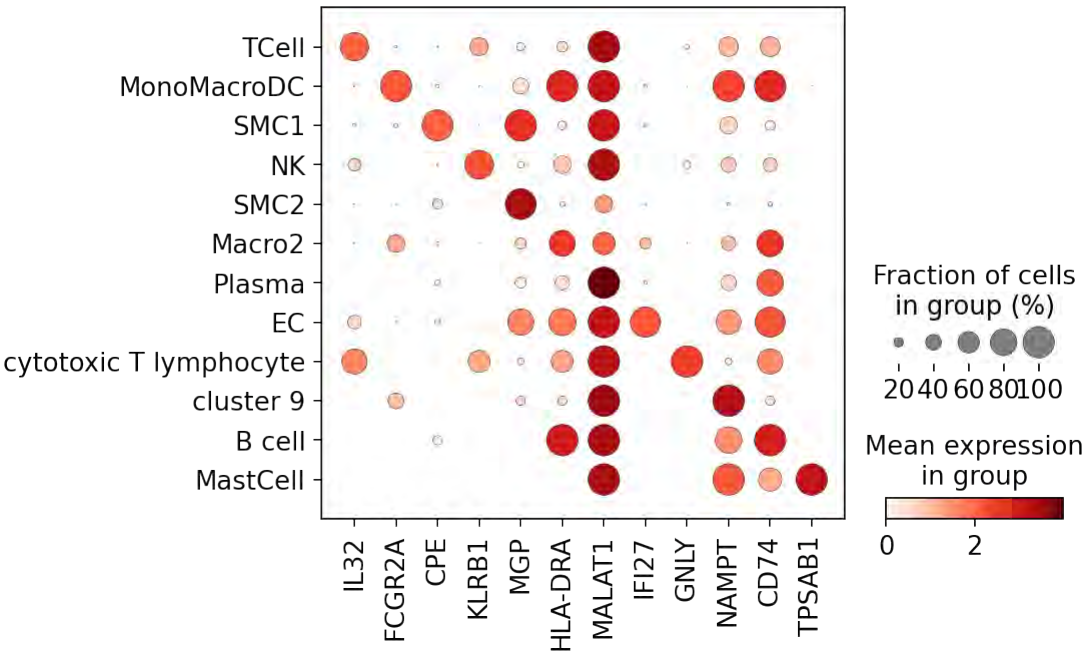
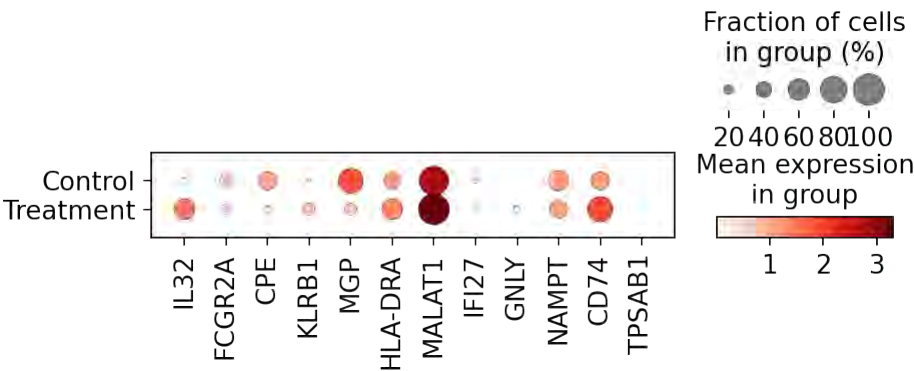


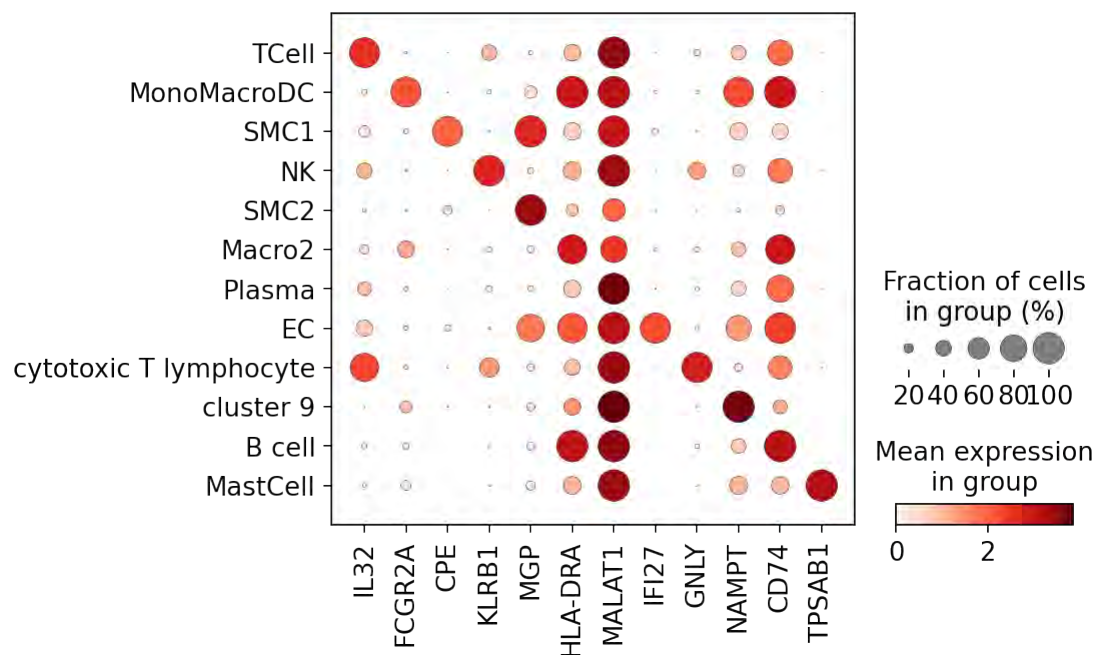
```
In [76]: #making a dotplot of the top marker genes
sc.pl.dotplot(adata, marker_genes, groupby='leiden');
```



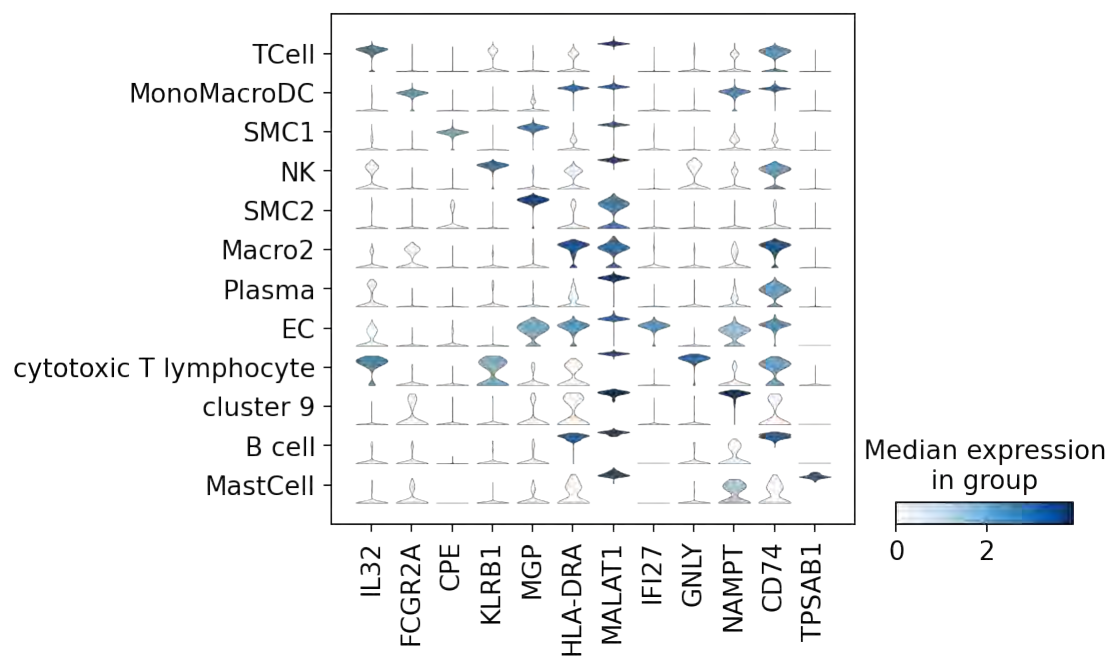
```
In [78]: #and comparing this dotplot by Treatment vs Control
sc.pl.dotplot(adata, marker_genes, groupby='Treatment');
```

```
sc.pl.dotplot(adata[adata.obs['Treatment'].isin(['Control'])], marker_genes, groupby='leiden')
sc.pl.dotplot(adata[adata.obs['Treatment'].isin(['Treatment'])], marker_genes, groupby='leiden')
```





```
In [79]: #and a very compact violin plot
sc.pl.stacked_violin(adata, marker_genes, groupby='leiden', rotation=90);
```



```
In [80]: #examine the adata object one more time
adata
```

```
Out[80]: AnnData object with n_obs × n_vars = 27224 × 21643
obs: 'Sample', 'Treatment', 'batch', 'n_genes', 'n_genes_by_counts', 'total_counts', 'total_counts_mt', 'pct_count'
```

```
s_mt', 'leiden'
var: 'mt', 'gene_ids', 'n_cells_by_counts', 'mean_counts', 'pct_dropout_by_counts', 'total_counts', 'highly_variable', 'means', 'dispersions', 'dispersions_norm', 'n_cells'
uns: 'Sample_colors', 'Treatment_colors', 'hvg', 'leiden', 'neighbors', 'pca', 'umap', 'log1p', 'leiden_colors', 'rank_genes_groups'
obsm: 'X_pca', 'X_umap'
```

Listing our output files

```
In [5]: #let's list our files that we've made:
!ls -lah
```

```
total 9.2G
drwxrwxr-x 4 welder-user users 4.0K Sep 22 21:02 .
drwxrwxr-x 4 welder-user users 4.0K Sep 21 15:10 ..
drwxrwxr-x 2 jupyter    users 4.0K Sep 21 15:12 data
-rw-rw-r-- 1 welder-user users  72 Sep 22 20:46 .delocalize.json
-rw-rw-r-- 1 welder-user users 15M Sep 22 21:02 human_aorta_cumulus_scanpy.ipynb
-rw-rw-r-- 1 jupyter    users 136 Sep 21 16:35 human_aorta_cumulus_scanpy-requirements.txt
drwxrwxr-x 2 jupyter    users 4.0K Sep 21 15:10 .ipynb_checkpoints
-rw-rw-r-- 1 jupyter    users 4.6G Sep 21 17:01 leiden_ttest_markers.txt
-rw-rw-r-- 1 jupyter    users 4.6G Sep 21 17:05 leiden_wilcoxon_markers.txt
```

```
In [ ]:
```